# The natural history of cervical HPV infection: unresolved issues

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Abstract | The identification of high-risk human papillomavirus (HPV) types as a necessary cause of cervical cancer offers the prospect of effective primary prevention and the possibility of improving the efficiency of cervical screening programmes. However, for these opportunities to be realized, a more complete understanding of the natural history of HPV infection, and its relationship to the development of epithelial abnormalities of the cervix, is required. We discuss areas of uncertainty, and their possible effect on disease prevention strategies.

## Adenocarcinoma

A malignant tumour originating in glandular tissue.

## Cross-sectional study

A study examining the association between disease and exposure at one point in time (a prevalence study). The temporal sequence of cause and effect cannot be determined with this study design.

## Episome

A piece of hereditary material that can exist as free, autonomously replicating DNA.

\*Cancer Research UK Institute for Cancer Studies, University of Birmingham, Edgbaston, Birmingham B15 2TT, UK. \*Cancer Research UK Clinical Trials Unit, University of Birmingham, Edgbaston, Birmingham B15 2TT, UK. Correspondence to C.B.J.W. e-mail: c.b.woodman@bham.ac.uk. doi:10.1038/nrc2050 Cervical cancer is the second most common cancer among women worldwide<sup>1</sup>. The majority of cases occur in the developing world, where, in most countries, it is the leading cause of cancer mortality in women<sup>2</sup>. In many developed countries, the incidence of squamous cell carcinoma of the cervix has been falling for some time, although that of adenocarcinoma of the cervix is now rising<sup>1,3,4</sup>.

Over 100 human papillomavirus (HPV) types have been identified, of which 40 infect the genital tract<sup>5</sup>. Cervical HPV infection is a common sexually transmitted infection. Most women are infected shortly after beginning their first sexual relationship<sup>6</sup>, with the highest prevalence seen in women under 25 years of age<sup>7,8</sup>. Thereafter, prevalence decreases rapidly. In young and middle-aged women, HPV infections are usually transient, at least when their duration is measured by how long the virus can be detected in cytological samples<sup>9-11</sup>. Virus might be detected only intermittently; and the concurrent or sequential detection of different HPV types is common<sup>12-18</sup>. Crosssectional studies indicate a second peak of infection in older women close to the age when the incidence of cervical cancer is maximum<sup>7,8</sup>.

A stream of epidemiological and laboratory-based research has identified infection with any one of 15 high-risk, or oncogenic, HPV types as a necessary but not sufficient cause of cervical cancer<sup>19–21</sup>. Whereas HPV18 is the type most strongly associated with adenocarcinoma of the cervix, HPV16, followed by HPV18, are the types most frequently detected when squamous cell carcinoma is diagnosed<sup>22,23</sup>. The frequency with which HPV16 is found in integrated forms increases with the severity of cervical neoplasia,

although in some women with invasive disease only episomal forms are detected. By contrast, HPV18 is almost always found in only integrated forms in women with high-grade cervical intraepithelial neoplasia (HGCIN) and invasive disease (FIG. 1). A bivalent HPV (types 16 and 18) and a quadrivalent HPV (types 6, 11, 16 and 18) vaccine are being evaluated in phase III clinical trials. Preliminary results indicate that these prophylactic HPV virus-like particle vaccines are effective in preventing infections with, and epithelial abnormalities caused by, the targeted HPV types<sup>20,24</sup>.

Cervical cancer is characterized by a well-defined pre-malignant phase that can be suspected on cytological examination of exfoliated cervical cells and confirmed on histological examination of cervical material. These pre-malignant changes represent a spectrum of histological abnormalities ranging from CIN1 (mild dysplasia) to CIN2 (moderate dysplasia) to CIN3 (severe dysplasia/carcinoma in situ). Although the treatment of cervical pre-malignant changes is therapeutically efficacious, it is also procedurally inefficient. This situation has arisen because of uncertainties surrounding the natural history of CIN. Cytological and histological examinations cannot reliably distinguish the few women with abnormal smears who will progress to invasive cancer from the vast majority of those whose abnormalities will spontaneously regress. Were a population-based prophylactic immunization programme introduced using either of the vaccines under consideration, and were it to achieve widespread coverage, then not only could it prevent up to 70% of all cervical cancers, but it could also reduce the costs of cervical screening programmes. However, it is unlikely that these screening programmes could be

# At a glance

- The most frequently detected human papillomavirus (HPV) type at the time of diagnosis of squamous cell carcinoma (SCC) is HPV16, followed by HPV18. HPV18 is the type most strongly associated with adenocarcinoma of the cervix, which is increasing in incidence at the same time as the incidence of SCC is falling.
- A bivalent HPV (types 16 and 18) and a quadrivalent HPV (types 6, 11, 16 and 18) vaccine are now being evaluated in phase III clinical trials, and have the potential to prevent about 70% of all cervical cancers. The quadrivalent HPV vaccine (Gardasil, Merck) has recently gained FDA and European Commission approval for use in women between the ages of 9–26.
- Although most women will at some time be infected with HPV, very few will progress to invasive disease. The identification of more robust markers of disease progression requires a more complete understanding of the natural history of type-specific HPV infections.
- It is unknown whether persistent HPV infections are characterized by the continuing detection of HPV, or by a state of viral latency during which the virus remains undetectable only to reappear later. A clearer understanding of these issues is essential for the effective implementation of screening strategies that include testing for HPV.
- Integration of HPV into the host genome results in a loss of negative-feedback control of oncogene expression, following disruption of the viral regulatory early gene E2.
   Whether the integration event itself is crucial to carcinogenesis is the subject of continuing debate.
- The prevalence of integrated forms varies with the infecting HPV type. Unlike HPV16, HPV18 integration seems virtually complete in women with cervical intraepithelial neoplasia grade 3 (CIN3) or invasive disease.
- The association between viral load and cervical disease varies with the HPV type, the physical state of the virus and the heterogeneity of the cervical lesion. The complexity of these relationships indicates that a measurement of viral load is not clinically useful.
- The concurrent or sequential detection of more than one HPV type is common. There
  is some evidence to indicate that the life cycles of different HPV types are not
  independent of each other, as has previously been assumed.
- HPV oncogenes can activate the cellular methylation machinery. The pattern of HPV gene methylation varies with the viral life cycle, the presence of disease and possibly the HPV type.
- Aberrant methylation of CpG islands in the promoter regions of tumour suppressor genes is one of several epigenetic changes that can contribute to carcinogenesis. The detection of these epigenetic changes in exfoliated cervical cells could improve the effectiveness of cervical screening programmes.

# Cervical intraepithelial neoplasia

(CIN) A disease characterized by precancerous changes in, and confined to, the epithelial cells lining the cervix.

#### Dysplasia

An epithelial abnormality in which the cells becomes disorganized, which is characterized by developmental changes in cell growth, shape and organization.

## Natural history

The course of disease or infection from onset to resolution.

#### Epigenetic

Inherited changes in gene expression resulting from altered chromatin structure or DNA modification rather than changes in DNA sequence. discontinued or even scaled-down for several reasons: immunization will only protect against HPV types that are targeted by the vaccine; protection will not be absolute and its longevity is uncertain; as yet, the possibility of genotype replacement cannot be excluded; and older women not covered by vaccination programmes will continue to be at risk.

Although most women will at some time have been infected with HPV, few will progress to invasive disease. Therefore, there is a continuing need for more robust markers of disease progression than those provided by morphological examination, or testing for the presence of high-risk HPV types. The identification of viral and host factors that modulate the risk of disease progression in women infected with HPV requires a more complete understanding of the natural history of HPV infection, and its relationship to the acquisition of epithelial abnormalities. Failure to do so runs the risk of compounding our imperfect understanding of the disease process with an even less perfect understanding of the natural history of HPV infection, therefore presenting the clinician with another set of management dilemmas.

The contribution of HPV to the pathogenesis of cervical cancer, and issues relating to vaccination have been discussed in previous reviews<sup>19,20</sup>. Here, we first consider the conceptual and methodological impediments to a clearer understanding of HPV as a persistent viral infection, and how this might affect the efficacy of HPV-based screening strategies. Before considering the possible synergy between different HPV types, we explore how the exposure-disease relationship varies with viral load, viral integration status and the infecting HPV type; these considerations are important in the context of prophylactic vaccines that might not achieve sterilizing immunity. Because progression to invasive disease requires an accumulation of genetic and epigenetic events, we consider how HPV can activate the cellular DNA methylation machinery and therefore epigenetically regulate both viral and cellular genes. Finally, we consider how the detection of these epigenetic changes in exfoliated cervical cells might improve the effectiveness of cervical screening programmes.

# **Persistent HPV infection**

Many viruses establish persistent infections that are characterized by continuous low or high levels of viral replication (for example human immunodeficiency virus and hepatitis B virus) or by periodic reactivation of a latent infection following apparently disease-free intervals (for example herpes simplex virus)<sup>25</sup> (FIG. 2).

Although it is now widely believed that a persistent infection with a high-risk HPV type is necessary for the development of HGCIN and invasive disease, the term 'persistence' has often been loosely defined when testing this hypothesis. In many of these studies, the occurrence of disease in women who test positive for HPV on two or more occasions (persistent infection) has been compared with that in women who test positive only once (transient infection). There are numerous conceptual problems with this approach. When defined in this way, the duration of a persistent infection is not a constant, but will vary depending on the interval between tests used in each study. In studies using this approach, the interval between tests ranged from 2 months to 7 years, with a median of 6 months<sup>12,26–49</sup>.

A more fundamental problem relates to inferences drawn from observations made at indeterminate points during the natural history of the infection. In these circumstances, the distinction between a persistent and transient infection is arbitrary to the extent that it is dependent on both the timing of the samples in relation to the natural history of the infection, and the interval between samples (FIG. 3a–c). In particular, it is impossible to determine how long a woman, who tests positive in her first sample, has been infected before that sample was taken (FIG. 3d). A more informative analysis is one that includes only women with incident HPV infections, for whom the date of onset of infection is readily available. However, even this study design does not guarantee clarity. For example, a study that included women with

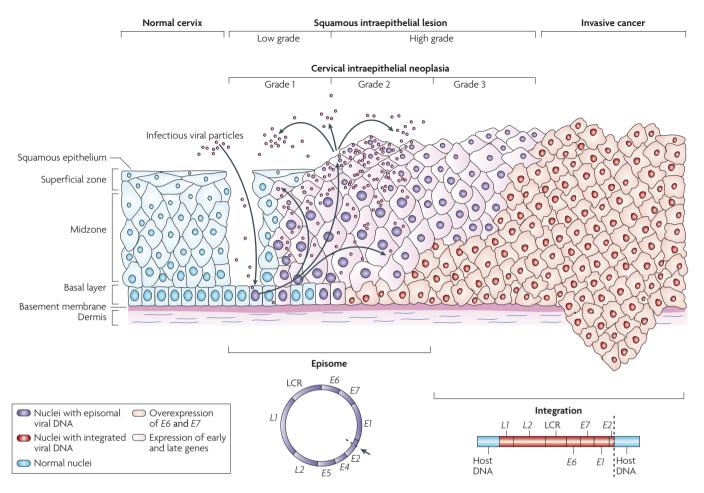


Figure 1 | **HPV-mediated progression to cervical cancer.** Basal cells in the cervical epithelium rest on the basement membrane, which is supported by the dermis. Human papillomavirus (HPV) is thought to access the basal cells through micro-abrasions in the cervical epithelium. Following infection, the early HPV genes *E1*, *E2*, *E4*, *E5*, *E6* and *E7* are expressed and the viral DNA replicates from episomal DNA (purple nuclei). In the upper layers of epithelium (the midzone and superficial zone) the viral genome is replicated further, and the late genes *L1* and *L2*, and *E4* are expressed. L1 and L2 encapsidate the viral genomes to form progeny virions in the nucleus. The shed virus can then initiate a new infection. Low-grade intraepithelial lesions support productive viral replication. An unknown number of high-risk HPV infections progress to high-grade cervical intraepithelial neoplasia (HGCIN). The progression of untreated lesions to microinvasive and invasive cancer is associated with the integration of the HPV genome into the host chromosomes (red nuclei), with associated loss or disruption of *E2*, and subsequent upregulation of *E6* and *E7* oncogene expression. LCR, long control region.

## Latent infection

The persistence of an infection in a host without symptoms and/or without being detectable.

#### Incident

A new occurrence of disease or infection in someone previously free of disease or infection. incident HPV infections defined a persistent infection as an infection of 6 months or more. This was less than the median duration of HPV infection in all cohort members, thereby classifying most women as having a persistent infection<sup>29</sup>. Although several investigators have reported the distribution of exposure times necessary for disease to occur<sup>9,50</sup>, some still characterize exposure levels according to the number of positive tests. This has led to apparently conflicting interpretations of the same data; the shorter the interval between tests, the more likely an infection will be defined as persistent<sup>26,30,32,51</sup> (FIG. 3e).

Many studies that have concluded that a persistent infection is a prerequisite for the development of cervical neoplasia can also be criticized on methodological grounds, because the sample taken at diagnosis is used to provide the second of the two consecutive positive samples that are necessary to define an infection as persistent<sup>30,32,33</sup> (FIG. 3f). Observations on exposure status made at or after the time of diagnosis are uninformative with respect to determining the exposure necessary for that disease to occur; an outcome cannot be attributed to a given level of exposure until that period of exposure has been completed<sup>52</sup>.

It is now clear that epithelial abnormalities of the cervix can be evident shortly after the first detection of HPV<sup>9,53</sup>. This is not to deny that HPV can establish a persistent infection, nor that a persistent infection is necessary for the development of invasive disease. However, it remains to be determined whether persistent infections are characterized by the continuing detection of HPV, or by a state of latency during which the virus remains undetectable only to reappear later. For example, one longitudinal study shows that HPV DNA can only

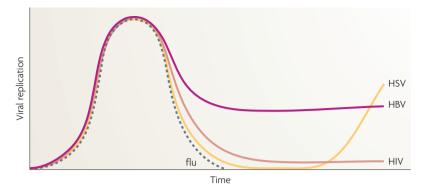


Figure 2 | **General patterns of infection.** To illustrate different patterns of persistent infection, the replication of herpes simplex virus (HSV), hepatitis B virus (HBV) and human immunodeficiency virus (HIV) is plotted as a function of time after infection. Acute infection (dashed grey line) is associated with clinical symptoms and the release of an infectious virus, such as influenza virus (flu). Persistent infection is associated with the production of an infectious virus, HBV for example (magenta line), for the lifetime of the host. Latent infection as seen in HSV infection (orange line) is a variation of persistent infection in which the acute infection is followed by a quiescent phase in which the virus productive cycle is absent or significantly reduced. The viral genome remains in a 'silent' state but can be intermittently reactivated into bouts of productive infection typified by long periods (years) between primary infection and the development of fatal symptoms: production of an infectious virus is either continuous at a low level or absent until failing immune control results in overwhelming virus production.

be detected in cytologically normal samples for a short time before cervical cancer is diagnosed<sup>32</sup>. A mechanism for latency has not been established so far, nor is it clear whether the differences between a latent and active cervical infection are qualitative or quantitative. However, a woman cannot be defined as having a persistent infection in any meaningful virological sense just because she tests positive for HPV on two occasions, some months apart, and therefore she should not, on the basis of this evidence, be considered to have a higher risk of cervical cancer. Nor can a woman, who tests positive for a specific HPV type, be considered to have cleared her infection when she first tests negative for that type. A clearer understanding of these issues is essential for the effective implementation of screening strategies, which include HPV testing.

# Longitudinal study

A study in which subjects are followed over a period of time so that the temporal sequence of potential cause and effect can be established.

# Triage

The classification of patients according to priority of need.

# Squamous intraepithelial lesion

(SIL). A disease characterized by the abnormal growth of squamous cells on the surface of the cervix. It is classified cytologically as low-grade (LSIL) or high-grade (HSIL) according to how much of the cervix is affected and how abnormal the cells are.

# Colposcopy

The visual examination of the uterine cervix with a magnifying lens to detect abnormal cells.

## HPV viral load

Among women who test positive for high-risk HPV types, cytological abnormality is more common in those with a high viral load<sup>49,54–58</sup>. The consistency of this finding, irrespective of the method used to measure viral load, persuaded many that the inclusion of such a measurement could improve the effectiveness of HPV-based screening and triage strategies. However, it is now clear that the relationship between viral load and disease is more complex than was previously thought. Whereas many cross-sectional studies reported an increase in viral load with increasing disease severity, others found either no association, or a higher viral load in women with low-grade squamous intraepithelial lesion (LSIL) than in those with high-grade squamous intraepithelial lesion (HSIL)<sup>49,54,55,57,58</sup> (TABLE 1;references 1–68 from Supplementary information S1 (table)). Longitudinal studies have also failed to find a consistent association between a baseline measurement of viral load and duration of infection, clearance of disease, and subsequent risk of acquisition or progression of disease<sup>33,42,59</sup>.

There are several possible explanations for these inconsistencies. Whereas the prevalence of integrated forms of HPV increases with increasing disease severity, integration itself is followed by a decrease in viral load. However, in almost all cross-sectional and longitudinal surveys that have measured viral load, integration status is undefined and might, of course, change over time<sup>60-63</sup>. In women with HGCIN, viral load is higher when low-grade CIN (LGCIN) is also present; in almost all studies, only the most severe histological abnormality is reported<sup>64</sup>. The acquisition of new HPV types is associated with both changes in viral load and with the development of new CIN lesions; therefore, measures of association might be unreliable in longitudinal studies that rely on a single baseline measurement of exposure<sup>51,59</sup>.

Finally, whereas many of the studies already cited used an assay that provided a measure of viral load aggregated across a panel of high-risk HPV types, it is now clear that the relationship between viral load and disease varies with HPV type. For example, cross-sectional studies show that HPV16 viral load increases with increasing disease severity, whereas that of HPV18 does not<sup>57,65,66</sup>. If the cytopathic effect observed in exfoliated cervical cells is a reflection of the viral load, then this might explain why the cytological changes detected after HPV18 infection underestimate the severity of the underlying histological abnormality, unlike those detected after HPV16 infection<sup>67</sup>. This is an important consideration because the benefits of cervical screening follow from the detection, investigation and treatment of epithelial abnormality, with the decision to refer for assessment by colposcopy usually based on the severity of the cytological abnormality. If HPV16 infection is more likely to be followed by a severe cytological abnormality than HPV18 infection, then screening programmes are more likely to interrupt the natural history of an HPV16 infection than that of an HPV18 infection. In many countries, screening programmes have not prevented an increase in the incidence of adenocarcinoma of the cervix, despite their success in preventing squamous cell carcinoma<sup>1,4,48,68</sup>. This failure has been attributed to the inaccessibility of these lesions to cytological sampling, but could also be explained in part by the strong association of adenocarcinoma with HPV18 infection, which in turn is associated with only minor cytological changes<sup>22,23,67,69</sup>. Although the initial optimism regarding the clinical value of HPV viral load testing now seems misplaced, robust measurements of type-specific viral load in samples in which the integration status is also defined, could provide useful insights into the natural history of HPV infections and their relationship to disease.

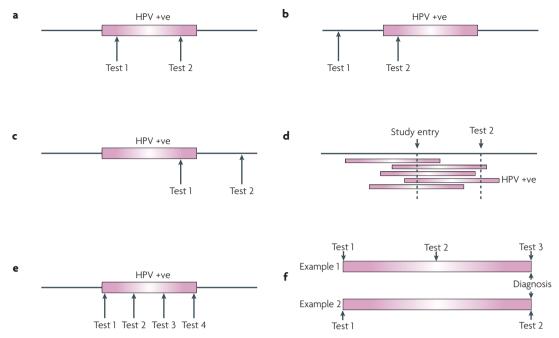


Figure 3 | **Persistent HPV infection.** The rectangles denote a period of time during which human papillomavirus (HPV) DNA sequences could be detected. In many studies, a woman is considered to have a persistent infection if she is HPV-DNA-positive in two or more consecutive tests, and a transient infection if she is positive only once. Panels **a**–**c** show how the same infection might be considered persistent or transient, depending on when the samples were taken. **a** | The infection is characterized as persistent. **b**, **c** | The same infection is now characterized as transient merely by changing the sampling times. **d** | These episodes of HPV infection are of identical duration, but began at different, undetermined times before entry to the study. Two of these infections would have been considered persistent, and three transient, on the basis of when the second test was performed. **e** | With a definition based on the number of positive tests, an infection is more likely to be considered persistent if a woman is tested more frequently. **f** | Example 1 fulfils the definition of a persistent infection, but the definition of a persistent infection was not fulfilled before the visit at which disease was detected, and therefore cannot be considered necessary for disease to occur.

# **HPV** integration

Human papillomavirus can be found in cervical material in episomal forms, integrated forms or in mixed forms that contain both. Viral integration into the host-cell genome occurs downstream of the early genes *E6* and *E7*, often in the *E1* or *E2* region; this disruption results in a loss of negative-feedback control of oncogene expression by the viral regulatory E2 protein (FIG. 1). Integrant-derived transcripts are more stable than those derived from episomal viral DNA, and HPV16 integration has been associated with a selective growth advantage for affected cells<sup>62,70</sup>.

The prevalence in exfoliated cervical cells or cervical tissue of episomal or integrated forms of HPV, or both, varies with the severity of disease, the infecting HPV type and the method used to determine the physical state of the virus<sup>60,61,71–115</sup>.

For example, HPV integration was once considered a late event in cervical carcinogenesis, because early studies rarely found integrated forms in women with CIN. However, when the physical state of the virus is determined by the failure to amplify full-length *E2* using PCR, the frequency of detection of integrated forms of HPV16 alone in women with CIN3 and in those with invasive disease is the same, and is comparable to that found in women with invasive disease using Southern blot hybridization (TABLE 2). Because the *E2* approach cannot distinguish integrated forms in the presence of episomal forms, the Southern blot detection method provides the most robust estimate of the overall prevalence of integrated forms. However, it might fail to detect integrated forms in the presence of a low viral load, which is more likely to be found following viral integration.

Integrated forms have also been detected in disease-free women and in those with LGCIN using *E2* PCR, although much less frequently. Unlike HPV16, HPV18 integration seems virtually complete in women with CIN3 or invasive disease, in whom episomal forms are rarely detected. Recently, realtime PCR assays, which simultaneously measure the HPV *E2* and *E6* copy numbers, have been used to determine the integration status. Although this assay seems ideally suited to exfoliated cervical cells and small cervical biopsy specimens, reconstitution experiments using different ratios of episomal and integrated forms of HPV16 show that, using this method, integrated forms can only be distinguished

Table 1   Association between high viral load and cervical neoplasia
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Positive association (number of studies)			
Yes	No		
4	4		
8	4		
3	2		
3	2		
29	1		
17 14 <sup>‡</sup> 6 <sup>‡</sup>	25 - -		
	Yes 4 8 3 3 29 17 14 <sup>‡</sup>		

\*Based on a single baseline measurement. <sup>‡</sup>In two studies, the exposure–disease relationship

varied with HPV type. This table was compiled from references 1–68 in Supplementary information S1. HPV, human papillomavirus; HSIL, high-grade squamous intraepithelial lesion; LSIL, low-grade squamous intraepithelial lesion.

in the presence of episomal forms when they are at least 100 times as common<sup>110</sup>. Furthermore, the size of the E2 product that can be detected using this method is constrained by the need to optimize amplification efficiency, and therefore it is not surprising that integrated forms alone are detected less frequently than when full-length E2 is amplified.

In almost all cervical cancers examined, integration is found at only one chromosomal site in the tumour cells, consistent with the idea that cervical cancer is a clonal disease<sup>116</sup>. Whether the integration event itself is crucial to carcinogenesis is the subject of ongoing debate. There are two schools of thought and both agree that, in women with invasive disease, integration preferentially occurs at common fragile sites; however, they disagree as to whether these sites are otherwise randomly distributed throughout the genome.

One school of thought argues that the overrepresentation of fragile sites is due to their greater susceptibility to integration-induced chromosomal alterations<sup>117-123</sup>; whereas the other argues that it is just a reflection of their greater accessibility<sup>124–126</sup>. One highlights the clustering of integration sites in certain chromosomal regions; whereas the other points to the absence of a specific cellular sequence motif in the transition sequence between the viral and cellular genome, or of recurrent integrations in a given region at the same locus. One emphasizes the number of cellular genes that are disrupted by HPV integration and that have been linked to carcinogenesis at other sites; whereas the other emphasizes the lack of evidence linking changes in expression of many of these genes to an integration event.

Although it is reasonable to suspect that integration will preferentially occur at those sites that confer a growth or survival advantage on the affected cells, the sites of HPV integration into the human genome have only been systematically studied in women with invasive cancer. There is no evidence that the location of integration sites in women who progressed to invasive disease differs from those who did not. So far, the most compelling evidence for insertional mutagenesis is provided by a report that describes transcriptional and protein overexpression of the proto-oncogene *MYC* in five cervical cell lines with HPV16 or HPV18 DNA sequences integrated at 8q24, but not in four cell lines in which integration occurred at other sites<sup>127</sup>. Integration at 8q24 has been found in approximately 10% of women with invasive disease and is reported to be more common in women with adenocarcinoma of the cervix and in those who test positive for HPV18 (REF. 120).

It has been proposed that the identification of integrated forms of HPV could be a useful biomarker for progressive disease. There are several problems with this proposal. First, the identification of small numbers of integrated forms in a background of mainly episomal forms will always be a technical challenge when only exfoliated cells are available for analysis. Second, if integrated genomes are often transcriptionally silent, or become so shortly after integration, then their detection might have limited prognostic usefulness. Although integrated forms are detected in over 40% of women with CIN3, active transcription of integrated forms has only been reported in 15% of such women<sup>93,128</sup> (TABLE 2). The detection of integrantderived transcripts could provide more useful prognostic information. However, it has been shown that cervical keratinocytes containing integrated forms will only emerge following a reduction in the number of E2-expressing episomes<sup>129</sup>. This loss of episomal E2, which is associated with the endogenous activation of anti-viral genes and is accelerated by exogenous interferon- $\beta$  (IFN $\beta$ ), increases the expression of viral oncogenes in cells containing integrated forms<sup>129-131</sup>. Therefore, testing for the absence of E2 in exfoliated cells, a technically much simpler proposition, merits evaluation as a marker of disease progression in HPV-positive women.

# Multiple HPV types: competition or synergy?

Longitudinal studies in both men and women show that the concurrent or sequential detection of more than one HPV type is common, and that this occurs more frequently than would be expected by chance<sup>12-15,18,132,133</sup>. If HPV types compete to colonize the cervical epithelium, then the prevalence of competing HPV types that are not targeted by vaccination could increase. Natural history studies offer no evidence of competition between HPV types, at least in so far as the risk of acquiring a new HPV type seems unrelated to a previous infection with another type<sup>14,15</sup>. However, publication of the postvaccination HPV-type-distributions in women who have already been vaccinated would provide more compelling evidence that this is the case. Natural history studies show an increased risk of acquisition of new HPV types in women already infected, compared with those who are HPV negative<sup>14,15</sup>. For example, the risk of acquiring HPV58 is up to seven times higher in women with an incident HPV16 or HPV18 infection compared with those who are not infected with these types<sup>18</sup>.

#### Fragile site

A site on a chromosome that tends to break more often than other sites.

#### Insertional mutagenesis

The occurrence of a mutation that is caused by the introduction of foreign DNA sequences into a gene.

#### Keratinocyte

An epidermal cell that produces the protein keratin.

Integrated forms alone			Episomal forms alone			Integrated ± episomal forms				
Full- length E1 PCR	Full- length E2 PCR	SB ± 2D GEL	E2:E6 ratio	ISH	SB ± 2D GEL	E2:E6 ratio	ISH	SB ± 2D GEL	E2:E6 ratio	ISH
28% (4; 267)	43% (14; 643)	49% (15; 469)	23% (3; 142)	67% (6; 147)	25% (15; 469)	17% (3; 142)	9% (6; 147)	66% (15;469)	70% (3; 142)	91% (6; 147)
-	42% (7; 212)	23% (4; 90)	13% (8; 243)	32% (2; 34)	56% (4; 90)	32% (8; 243)	12% (2; 34)	44% (4; 90)	62% (8; 243)	88% (2; 34)
-	0% (2; 44)	-	50% (2; 29)	-	-	15% (2; 29)	-	-	83% (2; 29)	-
-	5% (3;98)	0% (1; 12)	6% (3; 54)	-	100% (1; 12)	29% (3; 54)	-	0% (1; 12)	72% (3; 54)	-
-	11% (2; 37)	-	11% (1; 127)	-	-	20% (1; 127)	-	-	80% (1; 127)	-
-	100% (2;53)	98% (3; 51)	50% (1; 10)	97% (2; 31)	0% (3; 51)	0% (1; 10)	0% (2 (31)	100% (3; 51)	100% (1; 10)	100% (2; 31)
-	-	94% (1; 17)	-	-	0% (1; 17)	-	-	100% (1; 17)	-	-
	Integrate Full- length E1 PCR 28% (4; 267) - - - - - - - - - -	Integrated forms and         Full- length       Full- length         28%       43%         (4: 267)       (14: 643)         -       42%         (7; 212)       0%         -       0%         (2; 44)       5%         -       11%         -       11%         -       100%         -       100%	Integrate forms alore         Full- length E1 PCR       Full- length E2 PCR       SB $\pm$ 2D GEL         28%       43%       49%         (4; 267)       (14; 643)       (15; 469)         -       42%       23%         (7; 212)       (4; 90)       -         -       0%       -         (2; 44)       -       -         -       5%       0%         -       11%       -         -       100%       98%         -       100%       94%	Integrated forms alove           Full- length E1 PCR         Full- length E2 PCR         SB $\pm$ 2D GEL         E2:E6 ratio           28%         43%         49%         23%           (4; 267)         (14; 643)         (15; 469)         23%           -         42%         23%         13%           -         (2; 44)         -         50%           -         5%         0%         (1; 12)         6%           (2; 37)         -         11%         (1; 127)         11%           -         100%         98%         50%         (1; 10)           -         100%         94%         -         11%	Integrated forms alone           Full- length E1 PCR         SB $\pm$ 2D GEL         E2:E6 ratio         ISH           28% (4; 267)         43% (14; 643)         49% (15; 469)         23% (3; 142)         67% (6; 147)           -         42% (7; 212)         23% (4; 90)         13% (8; 243)         32% (2; 34)           -         0% (2; 44)         -         50% (2; 29)         -           -         5% (3; 98)         0% (1; 12)         6% (3; 54)         -           -         11% (2; 37)         -         11% (1; 127)         -           -         100% (2; 53)         98% (3; 51)         50% (1; 10)         97% (2; 31)           -         -         94%         -         -	Integrated forms alone       Episonal         Full- length E1 PCR       Full- length E2 PCR       SB $\pm$ 2D GEL       E2:E6 ratio       ISH       SB $\pm$ 2D GEL         28% (4; 267)       43% (14; 643)       49% (15; 469)       23% (3; 142)       67% (6; 147)       25% (15; 469)         -       42% (7; 212)       23% (4; 90)       13% (8; 243)       32% (2; 34)       56% (4; 90)         -       0% (2; 244)       -       50% (2; 29)       -       -         -       5% (3; 98)       0% (1; 12)       6% (3; 54)       -       100% (1; 12)         -       11% (2; 37)       -       11% (1; 127)       -       -         -       100% (2; 53)       98% (3; 51)       50% (1; 10)       97% (2; 31)       0% (3; 51)         -       -       94%       -       -       0%	Integrated forms alone         Episomal forms alone           Full- length E1 PCR         Full- length E2 PCR         SB $\pm$ 2D GEL         E2:E6 ratio         ISH         SB $\pm$ 2D GEL         E2:E6 ratio         ISH         SB $\pm$ 2D GEL         E2:E6 ratio           28% (4; 267)         43% (14; 643)         49% (15; 469)         23% (3; 142)         67% (6; 147)         25% (15; 469)         17% (3; 142)           -         42% (7; 212)         23% (4; 90)         67% (2; 243)         25% (2; 34)         17% (4; 90)         32% (8; 243)           -         0% (2; 44)         -         50% (2; 29)         -         -         15% (2; 29)           -         5% (3; 98)         0% (1; 12)         6% (3; 54)         -         100% (1; 12)         29% (1; 127)           -         11% (2; 37)         -         11% (1; 127)         -         -         20% (1; 127)           -         100% (2; 53)         98% (3; 51)         50% (1; 10)         97% (2; 31)         0% (3; 51)         0% (1; 10)           -         -         94%         -         -         0%         -	Integrated forms alone         Episomal forms alone           Full- length E1 PCR         Full- length E2 PCR         SB $\pm$ 2D GEL         F2:E6 ratio         ISH         SB $\pm$ 2D GEL         E2:E6 ratio         ISH           28% (4; 267)         43% (14; 643)         49% (15; 469)         23% (3; 142)         67% (6; 147)         25% (15; 469)         17% (3; 142)         9% (6; 147)           -         42% (7; 212)         23% (4; 90)         67% (8; 243)         25% (2; 34)         17% (3; 54)         9% (6; 147)           -         0% (7; 212)         -         50% (3; 98)         -         50% (2; 29)         -         15% (2; 24)         -           -         0% (2; 44)         -         50% (3; 98)         -         100% (3; 54)         -         -           -         11% (2; 37)         -         11% (1; 12)         -         -         20% (1; 12)         -           -         11% (2; 37)         -         11% (1; 12)         -         -         20% (1; 12)         -           -         100% (2; 53)         98% (3; 51)         50% (3; 51)         97% (1; 10)         0% (3; 51)         0% (3; 51)         0% (2 (31)           -         -         94%         -         -         0%	Integrated forms alone       Episomal forms alone       Integrate         Full- length E1 PCR       Full- length E2 PCR       SB $\pm$ 2D GEL       E2:E6 ratio       ISH       Episomal forms alone       Integrate         28% (4; 267)       43% (14; 643)       49% (15; 469)       23% (3; 142)       67% (6; 147)       25% (15; 469)       17% (3; 142)       9% (6; 147)       66% (15; 469)         -       42% (7; 212)       23% (4; 90)       13% (8; 243)       32% (2; 34)       56% (4; 90)       32% (8; 243)       12% (2; 29)       44% (4; 90)         -       0% (2; 44)       -       50% (1; 12)       -       15% (1; 12)       -       -         -       0% (3; 98)       0% (1; 12)       6%       -       100% (1; 12)       -       0% (1; 12)       -       -         -       11% (2; 37)       -       11% (1; 12)       -       -       20% (1; 12)       -       -         -       100% (2; 37)       98% (3; 51)       50% (1; 10)       97% (2; 31)       0% (3; 51)       0% (1; 10)       0% (2; 11)       -       -       -         -       100% (2; 53)       98% (3; 51)       50% (1; 10)       97% (2; 31)       0% (3; 51)       0% (3; 51)       0% (1; 10)       0% (2; 10) <td><math display="block"> \begin{array}{     larkgrated forms alone      larkgrated forms alo</math></td>	$ \begin{array}{     larkgrated forms alone      larkgrated forms alo$

# Table 2 | Frequency of HPV DNA forms detected according to HPV type, disease severity and detection method

Pooled analysis of studies reporting HPV16 or HPV18 integration status in cervical material. Studies were only included if they contributed 10 or more cases to at least one disease category. Median values are presented as a percentage followed by the number of studies contributing to the estimate, and the total number of cases in brackets. Episomal and integrated forms can be distinguished by the presence or absence of HPV-human DNA junction fragments using Southern blot (SB) analysis; by their different mobility using two-dimensional gel electrophoresis (2D gel); by their pattern of nuclear signalling using in *situ* hybridization (ISH); or by the *E2:E6* copy number ratio measured using real-time PCR. In real-time PCR, HPV is considered present in only episomal forms when *E2* and *E6* copy number are equivalent; in only integrated forms when *E2* is absent; and in mixed forms when *E2* is present but its copy number is less than that of *E6*. Failure to amplify full-length HPV *E1* or *E7* is consistent with the presence of integrated forms in the absence of episomal forms. This assay cannot identify integrated forms in the presence of episomal forms. CIN, cervical intraepithelial neoplasia; HPV, human papillomavirus; HSIL, high-grade squamous intraepithelial lesion.

The sequential detection of new HPV types could be the result of their sequential acquisition from different sexual partners. Whereas the first detection of HPV is clearly associated with the acquisition of a new sexual partner, the evidence linking the detection of new HPV types with a change in sexual partner in women who are already infected is not compelling<sup>18,134,135</sup>. Sequential transmission of several HPV types from the same partner cannot be excluded, but it seems unlikely. An alternative interpretation of these findings might be that more than one HPV type is transmitted simultaneously, and that their sequential detection after that is a consequence of replicative life cycles that are asynchronous and only occasionally overlapping. These life cycles might be interdependent. For example, in women with HSIL, HPV16 viral load is higher when other HPV types are present than when HPV16 is present alone<sup>136</sup>. In women with invasive disease, episomal forms of HPV16 are significantly more common in those who test positive for both HPV16 and HPV18 than in those who test positive for HPV16 alone<sup>85,137</sup>. If co-infection confers some mutual type-specific survival benefit, then the elimination of one HPV type could have an unexpected beneficial effect on the natural history of others.

## CpG island

A region of genomic DNA in which the frequency of the CG sequence is higher than in other regions.

# Epigenetic changes in cervical neoplasia

Both viral and host genes can be targeted by the cellular DNA methylation machinery. The pattern of methylation of HPV genes varies with the viral life cycle, the presence of disease and possibly the viral type<sup>138-141</sup>. The *de novo* methylation of HPV DNA could be a host defence mechanism for suppressing transcription of foreign DNA or a strategy that the virus uses to maintain a long-term infection, or both<sup>142,143</sup>. Treatment with the demethylating agent 5'-azacytidine has been shown to reactivate transcriptionally silent HPV integration sites in cervical cancer cell lines<sup>144</sup>. This raises the possibility, for which there is no empirical evidence so far, that the detection of HPV in older women might follow the epigenetic reactivation of previously silenced integration sites.

Aberrant methylation of CpG islands in the promoter regions of tumour suppressor genes (TSGs) is one of several epigenetic changes that can contribute to carcinogenesis<sup>145</sup>. TSG-promoter methylation varies qualitatively and quantitatively between Epstein-Barr virus (EBV)-positive and negative gastric cancers; hepatitis C-positive and negative hepatocellular carcinomas; and simian virus 40 (SV40)-positive and negative mesotheliomas146-157. It also varies between HPV-positive and negative vulval cancers, and between HPVpositive and negative head and neck cancers, but such comparisons are non-informative in cervical cancers, almost all of which are HPV-positive at diagnosis<sup>158-161</sup>. Cross-sectional comparisons have also failed to show HPV type-specific epigenetic changes, but have included only a small number of patients, and are likely to have been confounded by differences in histological type and disease severity.

Table 3   Epigenetic changes in cervical neoplasia									
No.	Gene	Disease-free controls	LG CIN	HG CIN	Squamous cell carcinoma	Cervical carcinoma	Adeno- carcinoma		
1	APC	18% (90; 4)	32% (37; 1)	34% (38; 1)	24% (238; 5)	32%* (88; 1)	54%* (65; 4)		
2	CCNA1	0% (25; 1)	0% (13; 1)	36%* (11; 1)		93%* (30; 1)			
3	CDH1	0% (53; 4)	7% (42; 3)	22%* (60; 3)	61%* (170(3)	47%* (135; 4)	33%* (57; 3)		
4	CDKN2A	3% (254; 7)	12%* (120; 6)	29%* (237; 9)	32%* (407; 8)	22%* (372; 7)	20%* (110; 7)		
5	DAPK1	1% (184; 5)	6% (69; 3)	30%* (88; 3)	64%* (299; 6)	52%* (180; 3)	39%* (89; 5)		
6	HIC1	2% (43; 3)	52%* (54; 2)	70%* (91; 3)	20%* (108; 2)	71%* (79; 1)	63%* (27; 2)		
7	IGSF4	0% (25; 3)	0% (29; 2)	39%* (31; 2)	58%* (52; 1)	65%* (23; 1)			
8	RARB	0% (47; 4)	5% (83; 3)	15%* (61; 3)	30%* (117; 3)	40%* (121; 3)	15%* (13; 2)		
9	ROBO1	0% (51; 1)	7% (62; 1)	8%* (48; 1)		46%* (119; 1)			
10	SLIT1	0% (40; 1)	0% (48; 1)	10%* (39; 1)		53%* (119; 1)			
11	SLIT2	0% (51; 1)	2% (62; 1)	25%* (48; 1)		64%* (119; 1)			
12	FANCF	0% (18; 1)		0% (37; 1)		30%* (91; 1)			
13	FHIT	0% (50; 4)	3% (76; 2)	2% (63; 2)	12%* (77; 1)	24%* (189; 4)	0% (5; 1)		
14	MGMT	3% (206; 6)	4% (93; 3)	7% (74; 3)	11%* (217; 4)	14%* (109; 2)	12%* (51; 3)		
15	PTEN	0% (11(1)	15% (27; 2)	0% (11(1)	58%* (62; 1)				
16	RASSF1	3% (29; 3)	3% (58; 3)	1% (73; 3)	17%* (299; 7)	5% (110; 3)	26%* (132; 6)		
17	SLIT3	0% (40; 1)	4% (48; 1)	2% (42; 1)		49%* (118; 1)			
18	TERT	0% (14; 1)	0% (13; 1)	0% (31; 1)		62%* (76; 2)	0% (9; 1)		
19	TIMP3	0% (8; 1)	0% (13; 1)	16% (31; 1)		11% (171; 3)	55%* (38; 2)		
20	C15orf48	0% (21; 1)				36%* (22; 1)			
21	MT1G	5% (21; 1)				55%* (22; 1)			
22	POU2F3	0% (7; 1)			41%* (32; 1)		36% (14; 1)		
23	SFRP1	5% (21; 1)				58%* (22; 1)			
24	SPARC	5% (21; 1)				91%* (22; 1)			
25	TFPI2	38% (21; 1)				82%* (22; 1)			
26	TNFRSF10C	0% (12; 1)				100%* (50; 1)			
27	HSPA2		0% (13; 1)	3% (31; 1)		73%* (11(1)			
28	SOCS1		0% (13; 1)	7% (31; 1)		50%* (11; 1)			
29	TWIST1		0% (23; 1)	14% (22; 1)		43%* (56; 1)			
30	SOCS2		23% (13; 1)	45% (31; 1)		64%* (11; 1)			
31	CDH13		5% (41; 2)	14% (63; 2)		46%* (89; 1)			

Pooled analysis reporting as a percentage the detection of specific methylated genes in cervical material. Listed are those genes for which methylated forms in tissue or exfoliated cells, taken from women with invasive disease, were significantly more frequent (P < 0.05) than in disease-free controls or when these were unavailable, women with low-grade cervical intraepithelial neoplasia (LGCIN). Genes numbered 1–11 are also more likely to be methylated in high-grade cervical intraepithelial neoplasia (HGCIN), whereas those numbered 12–19 are not. The presence of methylated forms in HGCIN and in disease-free control tissues has yet to be established for genes numbered 20–26 and 27–31, respectively. \*Estimates that are significantly different from the baseline. The table includes the number of cases contributing to each estimate and the number of studies from which they are derived in brackets. 'Cervical carcinoma' refers to cases of invasive disease with unspecified histological type. This table was compiled from references 69–110 in Supplementary information S1. *APC*, adenomatosis polyposis coli; *C15orf48*, chromosome 15 open reading frame 48; CCNA1, cyclin A1; CDH1, E-cadherin; CDH13, H-cadherin; CDKN2A, cyclin-dependent kinase inhibitor 2A; DAPK1, death-associated protein kinase 1; FANCF, Fanconi anaemia, complementation group F; FHIT, fragile histidine triad gene; HIC1, hypermethylated in cancer 1; HSPA2, heat-shock 70kDa protein 2; IGSF4, immunoglobulin superfamily, member 4; MGMT, O-6-methylguanine-DNA methyltransferase; MT1G, metallothionein 1G; POU2F3, POU domain, class 2, transcription factor 3; PTEN, phosphatase and tensin homologue, mutated in multiple advanced cancers 1; RARB, retinoic acid receptor β; RASSF1, Ras association (RaICDS/AF6) domain family 1; ROBO1, roundabout, axon guidance receptor, homologue 1; SFRP1, secreted frizzled-related protein 1; SLIT, slit homologue; SOCS, suppressor of cytokine signalling 1; SPARC, secreted protein, acidic, cysteine-rich (osteonectin); TERT, telomerase rev

The DNA methyltransferases (DNMT1, DNMT3A and DNMT3B) are responsible for the initiation and maintenance of methylation, and are overexpressed in several solid and haematological malignancies<sup>145</sup>. Gene expression profiling shows increased expression of DNMT1 in short-term primary cervical cancer cultures, compared with normal cervical keratinocyte cultures. Genomewide microarray-based comparative genomic hybridization shows an increase in DNMT3B copy number that is associated with changes in mRNA expression in all HPV-immortalized cell lines, and in most cervical cancers analysed<sup>162,163</sup>.

Viral oncogenes can induce TSG methylation following activation of DNA methyltransferases. For example, the TSG cadherin 1 (CDH1) is methylated following activation of DNA methyltransferases by the EBV oncogene latent membrane protein 1 (LMP1), and by the hepatitis B oncogene HBxAg (hepatitis B virus X antigen). DNMT1 is activated by LMP1 through the AP1-JNK (activated protein 1-JUN N-terminal kinase) signalling pathway<sup>164–168</sup>. Although there is no evidence so far for HPV-induced methylation of TSGs, HPV16 E7 has been shown to bind DNMT1, and to stimulate its enzymatic activity<sup>169</sup>. Moreover, similar to the BK virus large T antigen and the adenovirus E1A protein, E7 might also activate transcription of DNMT1. DNMT1 is a target of the transcription factor E2F1 and E7 can inactivate members of the pocket protein family that inhibit E2F family members. Therefore E7 could stimulate E2F activity and DNMT1 transcription<sup>170–172</sup>.

TABLE 3 lists those genes that are commonly found in methylated forms in exfoliated cervical cells or cervical tissue<sup>152</sup> (see also references 69–110 in Supplementary information S1). For some genes, the prevalence of methylated forms increases with disease severity; for others, methylated forms are only detected in women with invasive disease. For some of those genes, an association with HPV and cervical carcinogenesis has already been described. For example, *SFRP1* (secreted frizzled-related protein 1) is a negative regulator of the Wnt signalling pathway, the activation of which is required for the transformation of HPV-expressing human keratinocytes; and methylation of *SFRP4*, another negative regulator of this pathway, is associated with the detection of HPV16 in head and neck cancers<sup>161,173</sup>.

Other TSGs have been associated with HPV and carcinogenesis. SOCS1 (suppressor of cytokine signalling 1) interacts with HPV16 E7 protein and induces its ubiquitylation and degradation; PTEN (phosphatase and tensin homologue) inactivates STAT3 (signal transducer and activator of transcription 3) in HPV-infected cells; and POU2F3 (POU domain, class 2, transcription factor 3) might be involved in differentiation-dependent regulation of HPV transcription174-177. A screening test based on the detection of methylated forms of any one of these TSGs would lack sensitivity. However, the remarkable absence of methylated forms from disease-free women indicates that a test including more than one TSG, could improve sensitivity without the usual loss of specificity that follows the application of screening tests in parallel. Preliminary

studies using this strategy so far have confirmed the impression of high specificity without achieving the required sensitivity<sup>178,179</sup>.

# **Future directions**

A clearer understanding of HPV persistence is essential. Ultimately, cohort studies with long-term follow-up, using sensitive measures of type-specific viral load, will determine the incidence of true persistent HPV infections, the characteristics of that type of infection and its contribution to carcinogenesis. A more immediate goal would be to determine how often, in which epithelial cell subpopulations and in what form, HPV DNA sequences can be detected in cervical tissue after they can no longer be detected in cytological samples. Also of interest are age-related changes in the patterns of viral genome methylation, given the possibility that HPV persists in an epigenetically regulated latent state.

There has been a trend towards conflating all HPV types associated with cervical neoplasia, with a view to establishing whether testing for the presence of any one of a panel of high-risk types improves the effectiveness of cervical screening programmes. This has distracted attention from type-specific differences in the natural history of HPV infection that affect the exposure–disease relationship. For example, it will be important to determine for different HPV types how often and how soon viral integration follows an incident HPV infection; for how long following an integration event can integrant-derived transcripts be detected; and, once silenced, how often are transcription centres reactivated.

The contribution of TSG methylation to the initiation or progression of CIN has not been defined. Indeed, for no site of cancer has the risk of incident disease in disease-free individuals, or the risk of disease progression in those who already have pre-malignant changes, been related to the presence of TSG methylation in baseline material. Nor has the extent to which epigenetic changes explain the role of known carcinogens in the oncogenic process, been adequately explored. For example, the successful eradication from the stomach of another infectious agent, Helicobacter *pylori*, is followed by a reduction in the methylation density of CDH1, a TSG inactivated during gastric carcinogenesis<sup>180</sup>. Only longitudinal studies in women with incident HPV infections can distinguish those epigenetic events directly attributable to HPV infection or to co-factors such as smoking<sup>161</sup>, from those secondary to the disease process. Longitudinal studies in women with prevalent HPV infections will show how often epigenetic changes persist after HPV DNA sequences can no longer be detected, or after the successful treatment of CIN. The ease with which the cervix can be sampled, unlike almost all other sites of cancer, makes it an ideal model to explore how progression to a pre-malignant phenotype in vivo can be explained by genetic and epigenetic changes, and how these changes are linked to the acquisition of an oncogenic virus. The insights provided could be relevant to other models of viral carcinogenesis.

## DNA methyltransferase

An enzyme that transfers a methyl group to DNA. DNMT1 is the most abundant methyltransferase and is the main maintenance methyltransferase. DNMT3A and DNMT3B are the main *de novo* methyltransferases.

#### Pocket protein

The pocket protein family includes three proteins, RB (retinoblastoma), RBL1 (retinoblastoma-like 1) and RBL2. They have a crucial role in cell-cycle regulation through interaction with the E2F transcription factors.

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#### Competing interests statement

The authors declare competing financial interests: see Web version for details.

## DATABASES

The following terms in this article are linked online to: Entrez Gene:

http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=gene *CDH1* | DNMT1 | DNMT3A | DNMT3B | IFNβ | *LMP1* | *MYC* | POU2F3 | *PTEN* | *SFRP1* | *SFRP4* | *SOCS1* | *STAT3*  **National Cancer Institute:** http://www.cancer.gov Cervical cancer

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Division of Cancer Studies, University of Birmingham: http://www.cancerstudies.bham.ac.uk

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